

11-28-2023

## Genome Sequence and Annotation of the B3 Mycobacteriophage Phayeta

Emily Bishop  
*Coastal Carolina University*

Warren Earley  
*Coastal Carolina University*

Alexandra Greco  
*Coastal Carolina University*

Emma Hofseth  
*Coastal Carolina University*

Emma Kinerson  
*Coastal Carolina University*

*See next page for additional authors*

Follow this and additional works at: <https://digitalcommons.coastal.edu/biology>

---

### Recommended Citation

Bishop, E., Earley, W., Greco, A., Hofseth, E., Kinerson, E., Lafayette, B., Llanot-Arocho, N., Mazen, B., Cevasco, M., & Williams, D.C. (2023). Genome sequence and annotation of the B3 mycobacteriophage Phayeta. *Microbiology Resource Announcements*, e00915-23. <https://doi.org/10.1128/MRA.00915-23>. Available at <https://digitalcommons.coastal.edu/biology/6/>.

This Article is brought to you for free and open access by the College of Science at CCU Digital Commons. It has been accepted for inclusion in Biology by an authorized administrator of CCU Digital Commons. For more information, please contact [commons@coastal.edu](mailto:commons@coastal.edu).

---

**Authors**

Emily Bishop, Warren Earley, Alexandra Greco, Emma Hofseth, Emma Kinerson, Brandon Lafayette, Nestor Llanot-Arocho, Brittney Mazon, Megan Cevalco, and Daniel C. Williams

# Genome sequence and annotation of the B3 mycobacteriophage Phayeta

Emily Bishop,<sup>1</sup> Warren Earley,<sup>1</sup> Alexandra Greco,<sup>1</sup> Emma Hofseth,<sup>1</sup> Emma Kinerson,<sup>1</sup> Brandon Lafayette,<sup>1</sup> Nestor Llanot-Arocho,<sup>1</sup> Brittney Mazen,<sup>1</sup> Megan Cevasco,<sup>1</sup> Daniel C. Williams<sup>1</sup>

**AUTHOR AFFILIATION** See affiliation list on p. 2.

**ABSTRACT** Mycobacteriophage Phayeta was extracted from soil near Myrtle Beach, South Carolina using *Mycobacterium smegmatis* as a host. Annotation of the 68,700 base-pair circularly permuted genome identified 104 predicted protein-encoding genes, 34 of which have functional assignments.

**KEYWORDS** Bacteriophage

*Mycobacterium smegmatis* is a genetically tractable model of *Mycobacterium tuberculosis* and phages that infect *M. smegmatis* have the potential to be therapeutically useful (1, 2). Mycobacteriophage Phayeta was directly isolated from a soil sample taken near Myrtle Beach, South Carolina (Global Positioning Coordinates 33.75493N, 78.90047 W), using standard protocols (3). The soil sample was washed in 7H9 liquid media, filtered (0.22  $\mu$ m), and 500  $\mu$ L of the filtrate mixed with 250 mL of saturated *M. smegmatis* mc<sup>2</sup>155 for 15 minutes before being plated in top agar and incubated at 37°C for 48 hours. Phayeta produced small clear plaques that are characteristic of non-temperate phages (Fig. 1A). Plaque size was measured with *ImageJ* and ranged from 0.5 mm to 1.7 mm with an average of  $1.0 \pm 0.2$  mm (mean  $\pm$  SD,  $n = 50$ ) (4, 5). Negative-staining transmission electron microscopy of Phayeta (Fig. 1B) revealed a siphovirus morphology consisting of an isometric capsid (diameter =  $76.6 \pm 3.6$  nm) and a long flexible tail ( $274.7 \pm 11.6$ ,  $n = 9$  viral particles).

Phayeta DNA was extracted from high-titer lysates using the Wizard DNA Clean-Up Kit (Promega) and analyzed by agarose gel electrophoresis. Intact DNA was prepared with the Ultra II Library Kit (New England Biolabs) and sequenced on the Illumina MiSeq platform (v3 reagents). A total of 446,915 single-end 150 base reads provided 278 $\times$  coverage of the Phayeta genome. Sequence reads were assembled and verified using Newbler (v2.9) and Consed (v29) (6). The genome is 68,700 base-pairs long, has 67.5% GC content, and contains circularly permuted genomic ends. Comparison of gene nucleotide similarity of phages within the Actinobacteriophage Database ([phagesdb.org](http://phagesdb.org)), placed Phayeta in subcluster B3 (7). Among this cluster, Phayeta shares the greatest gene content similarity with Casbah (94.6%) and Kronus (91.7%).

Initial draft annotation of Phayeta was done using the GenMark (v2.5), Glimmer (v3.02), and ARAGORN algorithms within DNA Master (v5.23.56) using the bacterial, archaeal, and plant plastid code (8–10). Subsequent manual annotations employed Phamerator (v7), *Starterator* (v1.3), and *PECAAN* (11). Both PhagesDB and NCBI non-redundant databases were searched for sequence-based homology comparison via BLAST (7, 12). Predicted protein structure comparisons used HHPRED (v3.18) to search the PDB mmCIF70, Pfam-A, and NCBI Conserved Domain databases using default parameters. A total of 104 protein-encoding genes were identified, 34 of which have a functional assignment.

**Editor** John J. Dennehy, Queens College, Queens, New York, USA

Address correspondence to Daniel C. Williams, [dwilliams@coastal.edu](mailto:dwilliams@coastal.edu).

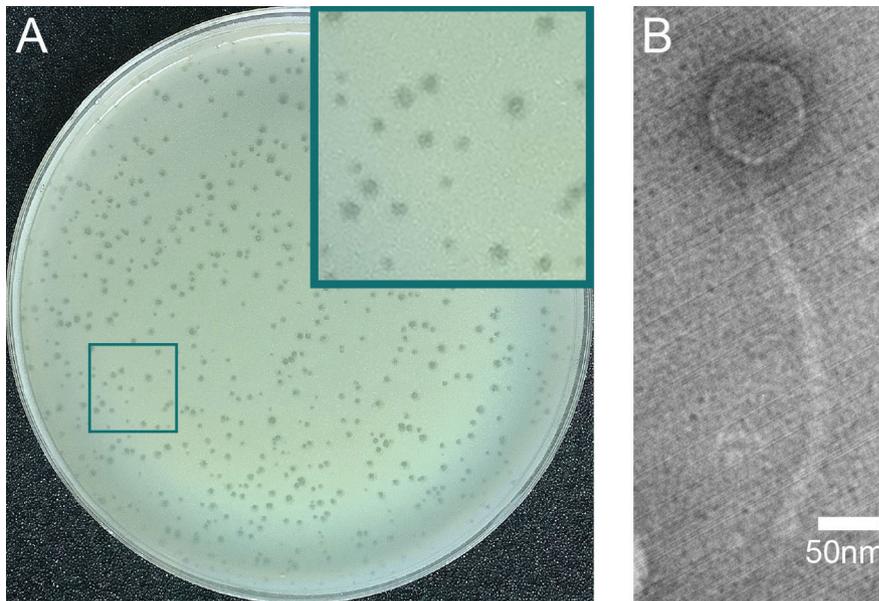
The authors declare no conflict of interest.

**Received** 28 September 2023

**Accepted** 29 September 2023

**Published** 28 November 2023

Copyright © 2023 Bishop et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).



**FIG 1** The mycobacteriophage Phayeta. (A) Plaque assay. Purified samples of Phayeta (10  $\mu$ l) were mixed with 250  $\mu$ l of saturated *M. smegmatis* MC<sup>2</sup>155, then combined with 3 mL of molten 7H9 top-agar and poured on 90 mm plates. Incubation at 37°C for 48 hours resulted in the production of clear small plaques. (B) Transmission electron microscopy of Phayeta negatively stained with 1% uranyl acetate and imaged using a JEOL JEM 1230 at 100 kV.

Most genes on the left arm of Phayeta encode structural or assembly proteins with the right arm containing genes involved in DNA replication. Consistent with a non-temperate lifestyle, Phayeta lacks genes encoding an identifiable integrase or repressor. Both Lysin A and Lysin B are present. Although homology searching failed to identify a holin-encoding gene, a small cluster of transmembrane-segment-containing genes (gp17-gp20) have sequence and syntenic similarities with genes of *Gordonia* phages that are a predicted holin cassette (13). There are five minor tail proteins, including gp33 which shows full-length similarity to Corofin gp33. Both these phages have a single nucleotide indel polymorphism in this gene but at different locations suggesting distinct mutational events.

#### ACKNOWLEDGMENTS

We thank Dan Russell and Ching-Chung Ko of the Pittsburgh Bacteriophage Institute for sequencing and genome assembly, Julian Smith III and Victoria Frost of Winthrop University for their electron microscopy service, and the Howard Hughes Medical Institute SEA-PHAGES program and Coastal Carolina University Biology Department for support.

#### AUTHOR AFFILIATION

<sup>1</sup>Department of Biology, Coastal Carolina University, Conway, South Carolina, USA

#### AUTHOR ORCID<sub>s</sub>

Megan Cevasco  <http://orcid.org/0000-0003-2027-2225>

Daniel C. Williams  <http://orcid.org/0000-0002-7900-480X>

#### DATA AVAILABILITY

Phayeta is available at GenBank with Accession No. [OR159655](https://www.ncbi.nlm.nih.gov/nuclseq/OR159655) and Sequence Read Archive No. [SRX20630268](https://www.ncbi.nlm.nih.gov/sra/SRX20630268).

## REFERENCES

1. Shiloh MU, Champion PAD. 2010. To catch a killer. what can mycobacterial models teach us about *Mycobacterium tuberculosis* pathogenesis? *Curr Opin Microbiol* 13:86–92. <https://doi.org/10.1016/j.mib.2009.11.006>
2. Hatfull GF. 2022. Mycobacteriophages: from petri dish to patient. *PLoS Pathog* 18:e1010602. <https://doi.org/10.1371/journal.ppat.1010602>
3. Poxleitner M, Pope WH, Jacobs-Sera D, Sivanathan V, Hatfull GF. 2018. SEA-PHAGES phage discovery guide. Available from: <https://seaphage-sphagediscoveryguide.helpdocsonline.com/home>
4. Collins TJ. 2007. ImageJ for microscopy. *Biotechniques* 43:25–30. <https://doi.org/10.2144/000112517>
5. Schindelin J, Arganda-Carreras I, Frise E, Kaynig V, Longair M, Pietzsch T, Preibisch S, Rueden C, Saalfeld S, Schmid B, Tinevez J-Y, White DJ, Hartenstein V, Eliceiri K, Tomancak P, Cardona A. 2012. Fiji: an open-source platform for biological-image analysis. *Nat Methods* 9:676–682. <https://doi.org/10.1038/nmeth.2019>
6. Russell DA. 2018. Sequencing, assembling, and finishing complete bacteriophage genomes, p 109–125. In Clokie MRJ, AM Kropinski, R Lavigne (ed), *Bacteriophages*. Springer, New York, New York, NY. <https://doi.org/10.1007/978-1-4939-7343-9>
7. Russell DA, Hatfull GF. 2017. PhagesDB: the actinobacteriophage database. *Bioinformatics* 33:784–786. <https://doi.org/10.1093/bioinformatics/btw711>
8. Besemer J, Borodovsky M. 2005. GeneMark: web software for gene finding in prokaryotes, eukaryotes and viruses. *Nucleic Acids Res* 33:W451–W454. <https://doi.org/10.1093/nar/gki487>
9. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with glimmer. *Bioinformatics* 23:673–679. <https://doi.org/10.1093/bioinformatics/btm009>
10. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>
11. Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. *BMC Bioinformatics* 12:395. <https://doi.org/10.1186/1471-2105-12-395>
12. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2)
13. Pollenz RS, Bland J, Pope WH. 2022. Bioinformatic characterization of endolysins and holin-like membrane proteins in the lysis cassette of phages that infect *Gordonia rubripertincta*. *PLoS One* 17:e0276603. <https://doi.org/10.1371/journal.pone.0276603>